

Taxonomy & Inventories

Disclisioprocta edmondsii (Butler, 1882) comb. nov. (Lepidoptera, Geometridae, Larentiinae)

Héctor A. Vargas ‡

‡ Universidad de Tarapacá, Facultad de Ciencias Agronómicas, Departamento de Recursos Ambientales, Arica, Chile

Corresponding author: Héctor A. Vargas (lepvargas@gmail.com)

Academic editor: Shinichi Nakahara

Received: 16 Dec 2022 | Accepted: 04 Jan 2023 | Published: 11 Jan 2023

Citation: Vargas HA (2023) Disclisioprocta edmondsii (Butler, 1882) comb. nov. (Lepidoptera, Geometridae,

Larentiinae). Biodiversity Data Journal 11: e98935. https://doi.org/10.3897/BDJ.11.e98935

Abstract

Background

The generic assignment of the geometrid moth *Xanthorhoe edmondsii* (Butler, 1882) (Lepidoptera, Geometridae, Larentiinae), originally described under *Hypochroma* Guenée, [1858], a junior homonym of *Hypochroma* Herrich-Schäffer, [1855] (Geometridae, Ennominae), is assessed using genitalia morphology and analysis of mitochondrial DNA sequences.

New information

Morphological characters revealed closeness to the type species of *Disclisioprocta* Wallengren, 1861 (Larentiinae). In agreement with morphology, the molecular analysis clustered *X. edmondsii* with species of *Disclisioprocta* in a well-supported monophyletic group distantly related to members of *Xanthorhoe* Hübner, [1825]. Accordingly, *Disclisioprocta edmondsii* (Butler, 1882) comb. nov. is proposed.

Keywords

DNA barcoding, genitalia morphology, geometrid moths, Neotropical

Introduction

Natural environments of South America harbour a high diversity of geometrid moths (Lepidoptera, Geometridae), whose taxonomy remains insufficiently studied (Hausmann and Parra 2009, Brehm et al. 2016, Murillo-Ramos et al. 2021). Besides the frequent discovery of new species (Brehm 2018, Ramos-González et al. 2019, Moraes et al. 2021, Vargas 2021), generic assignments of most of the already described species deserve assessment, as suggested by new combinations arising in taxonomic revisions (Parra 2018) and molecular phylogenetic analyses (Brehm et al. 2019).

The geometrid moth *Xanthorhoe edmondsii* (Butler, 1882) (Geometridae, Larentiinae) is known from Chile and Argentina (Butler 1882, Chalup 2014). It was originally described under *Hypochroma* Guenée, [1858], a junior homonym of *Hypochroma* Herrich-Schäffer, [1855] (Geometridae, Ennominae) (Parsons et al. 1999). Individuals from northern Chile reared from larvae collected on the ornamental plant *Bougainvillea glabra* Choisy (Nyctaginaceae) were misidentified as *Chrismopteryx undularia* (Blanchard, 1852), based on comparison with material from central Chile deposited in the Museo Nacional de Historia Natural de Santiago (Vargas et al. 2010). Subsequent comparison with a photo of the type material deposited in the Natural History Museum, London, UK, allowed concluding that the specimens examined by Vargas et al. (2010) belong to *X. edmondsii*.

Morphological characters of the genitalia of geometrid moths are extremely useful in generic assignments (Pitkin 2002, Parra 2018, Viidalepp and Lindt 2019), which can be reinforced by phylogenetic analysis of DNA sequences (Brehm et al. 2019, Matson 2022, Wanke et al. 2022). An examination of the genitalia of *X. edmondsii* revealed remarkable morphological differences with *Xanthorhoe montanata* ([Denis & Schiffermüller], 1775), the type species of *Xanthorhoe* Hübner, [1825], suggesting instead closeness with *Disclisioprocta stellata* (Guenée, [1858]), the type species of *Disclisioprocta* Wallengren, 1861. The aim of this study is to propose a new generic assignment for *X. edmondsii*, based on genitalia morphology and analysis of mitochondrial DNA sequences.

Materials and methods

Specimens examined in this study were collected at light or reared from larvae collected on *B. glabra* in the Azapa Valley (18°31'16"S, 70°10'42"W), Arica Province, northern Chile. Photos of the genitalia were taken with a Leica Flexacam C1 digital camera attached to a Leica M125 stereomicroscope. Each image was constructed with about 5–10 photos assembled with the software Helicon Focus 8. The specimens of *X. edmondsii* are deposited in the "Colección Entomológica de la Universidad de Tarapacá" (IDEA), Arica,

Chile. The specimens of *D. stellata* are deposited in the "Coleção Pe. Jesus de Santiago Moure (DZUP)", Universidade Federal do Paraná, Curitiba, Paraná, Brazil.

Genomic DNA was extracted from two legs of a male adult using the QIAamp Fast DNA Tissue Kit, following the manufacturer's instructions. DNA purification, PCR amplification and sequencing of the barcode region (Hebert et al. 2003) with the primers LCO1490 and HCO2198 (Folmer et al. 1994) were performed at Macrogen Inc. (Seoul, South Korea). The PCR programme was 5 min at 94°C, 35 cycles of 30 s at 94°C, 30 s at 47°C, 1 min at 72°C and a final elongation step of 10 min at 72°C. Analysis of this mitochondrial marker represents a helpful tool in generic assignments of Lepidoptera, including Geometridae (Wanke et al. 2019, Wanke et al. 2022, Vargas et al. 2022). The sequence of X. edmondsii was submitted to a Maximum Likelihood (ML) analysis with additional representatives of Larentiinae downloaded from BOLD (Ratnasingham and Hebert 2007), following the classification provided by Brehm et al. (2019) in which Disclisioprocta belongs to an unnamed clade sister to Euphyiini. The alignment included sequences of Euphyia Hübner, [1825] and Oligopleura Herrich-Schäffer, [1855] as representatives of this tribe, sequences of Xanthorhoe due to the current generic adscription of X. edmondsii (Parsons et al. 1999) and sequences of Scotopteryx Hübner, [1825] as outgroups. The software MEGA11 (Tamura et al. 2021) was used to perform sequence alignment with the ClustalW method and to determine genetic distance using the Kimura 2-Parameter (K2P) method. Before the ML analysis, the substitution saturation of the alignment was assessed with the Xia test, using the software DAMBE7 (Xia 2018). The ML analysis was performed using the software IQTREE 1.6.12 (Nguyen et al. 2014) in the web interface W-IQ-TREE (Trifinopoulos et al. 2016) with data partitioned to codon position. ModelFinder (Kalyaanamoorthy et al. 2017) selected TNe+I, F81+F and TN+F+G4 as the best fit models for 1st, 2nd and 3rd partitions, respectively. Branch support was assessed with 1000 replications of the Shimodaira-Hasegawa-like approximate likelihood ratio test (SH-aLRT, Guindon et al. (2010)) and ultrafast bootstrap (UFBoot, Hoang et al. (2017)). The unrooted tree was visualised in FigTree (Rambaut 2014) to root on Scotopteryx.

Taxon treatment

Disclisioprocta edmondsii (Butler, 1882) comb. nov.

Nomenclature

Hypochroma edmondsii Butler, 1882, p. 364. Angulo and Casanueva (1981), p. 21.

Xanthorhoe edmondsii (Butler, 1882): Parsons et al. (1999), p. 964.

Chrismopteryx undularia (Blanchard, 1852): Vargas et al. (2010), misidentification.

Materials

a. scientificName: Disclisioprocta edmondsii (Butler, 1882); higherClassification:
 Insecta;Lepidoptera;Geometridae;Larentiinae; continent: South America; country: Chile;

stateProvince: Arica; locality: Azapa Valley; decimalLatitude: -18.52; decimalLongitude: -70.18; samplingProtocol: Two males, three females emerged February 2006, reared fom larvae collected on *Bougainvillea glabra* January 2006; individualCount: 5; identifiedBy: Héctor A. Vargas; identificationRemarks: Genitalia slides HAV-1281, 1284, 1286, 1583, 1584; type: PhysicalObject; language: en; institutionCode: "Colección Entomológica de la Universidad de Tarapacá" (IDEA); basisOfRecord: "PreservedSpecimen"; occurrenceID: 99F2E982-FCB8-5FF0-B368-3F12C8EFF0A6

- b. scientificName: Disclisioprocta edmondsii (Butler, 1882); higherClassification:
 Insecta;Lepidoptera;Geometridae;Larentiinae; continent: South America; country: Chile;
 stateProvince: Arica; locality: Azapa Valley; decimalLatitude: -18.52; decimalLongitude:
 -70.18; samplingProtocol: Two males, one female September 2006 at light;
 individualCount: 3; identifiedBy: Héctor A. Vargas; identificationRemarks: Genitalia slides
 HAV-1283, 1285, 1287; type: PhysicalObject; language: en; institutionCode: "Colección
 Entomológica de la Universidad de Tarapacá" (IDEA); basisOfRecord:
 "PreservedSpecimen"; occurrenceID: 591A91A6-F832-5EFC-85C4-F03775D0D824
- c. scientificName: Disclisioprocta edmondsii (Butler, 1882); higherClassification: Insecta;Lepidoptera;Geometridae;Larentiinae; continent: South America; country: Chile; stateProvince: Arica; locality: Azapa Valley; decimalLatitude: -18.52; decimalLongitude: -70.18; samplingProtocol: One male May 2022 at light; individualCount: 1; associatedSequences: BOLD Process ID GEONC001-22; identifiedBy: Héctor A. Vargas; identificationRemarks: Genitalia slide HAV-1580; type: PhysicalObject; language: en; institutionCode: "Colección Entomológica de la Universidad de Tarapacá" (IDEA); basisOfRecord: "PreservedSpecimen"; occurrenceID: 6C800146-205F-5AA7-85A5-4470D74FCA88

Description

Male habitus in Fig. 1. Although the male abdominal segments VII and VIII are not part of the genitalia, these are described here and illustrated because the morphology of the sclerites of these segments can be modified in different groups of Larentiinae (Viidalepp 2011).



Figure 1. doi

Male adult of *Disclisioprocta edmondsii* (Butler, 1882) comb. nov., dorsal view. Scale bar 10 mm.

Male abdominal segments VII and VIII (Fig. 2). Segment VII mostly membranous; tergum a transverse stripe strongly posteriorly folded in the middle; sternum a transverse stripe; pleura with pair of coremata. Segment VIII mostly membranous; tergum an anterior transverse stripe with semicircular expansion on tips, connected by a short longitudinal stripe with a posterior rectangular transverse plate; sternum an anterior transverse stripe posteriorly curved in the middle, projected as a narrow longitudinal stripe posteriorly bifid, triangular expansion near tip of the anterior transverse stripe.

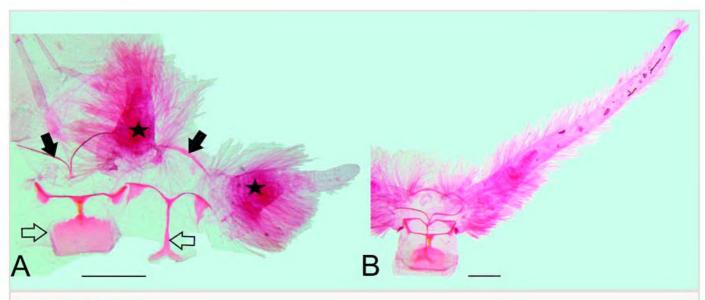


Figure 2. doi

Male abdominal segments VII and VIII of *Disclisioprocta edmondsii* (Butler, 1882) comb. nov. A) Coremata (stars) of segment VII and sclerites of segments VII (closed arrows) and VIII (open arrows); terga on left, sterna on right. B) Segments VII and VIII showing right corema of segment VII expanded. Scale bar 1 mm.

Male genitalia (Fig. 3). Uncus bifid with broad posterior concavity in the middle, truncate points slightly down-curved. Saccus with small rounded anterior projection. Subscaphium slightly sclerotised. Labides with lobe-like tip bearing setae. Manica heavily spinose. Juxta trapezoidal, ventral half of lateral margin broadly concave, ventral margin broadly concave. Valva elongated; costal sclerotised band not reaching apex; cucullus mostly membranous on distal half with abundant setae; sacculus broad, well-sclerotised; sacculus projection stout, apex almost reaches that of the distal margin of the cucullus, with a broader, dorsally projected basal process. Phallus cylindrical, anterior half straight, posterior half curved, with a small spine-like projection ventrally on posterior tip; vesica mostly membranous with a plate-like cornutus.

Female genitalia (Fig. 3). Papillae anales membranous, lobe-like, fused dorsally, posterior edge with setae on dorsal and lateral parts and elongated, flattened scales on ventral part. Apophyses posteriores rod-shaped, narrow, slightly longer than apophyses anteriores. Antrum well-sclerotised, flattened, ventrally curved in the middle, progressively straightening anteriorly. Ductus bursae membranous, about 2/3 length of the antrum. Corpus bursae membranous, spherical, with 5–7 stout spine-like signa arising ventrally from the anterior margin of a semicircular slightly sclerotised plate. Ductus seminalis arising near the posterior tip of ductus bursae.

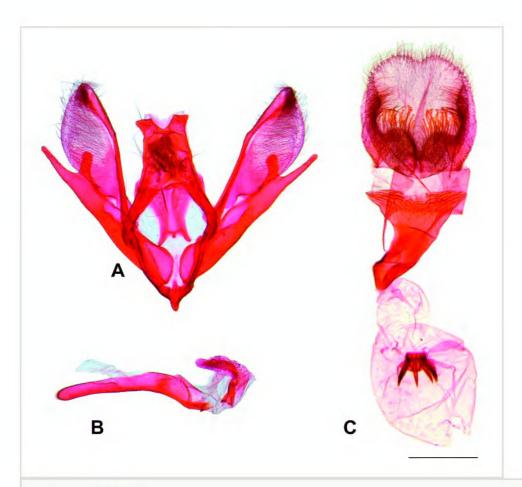


Figure 3. doi

Genitalia of *Disclisioprocta edmondsii* (Butler, 1882) comb. nov. A) Male genitalia, phallus removed. B) Phallus. C) Female genitalia. Scale bar 1 mm.

Molecular analysis

Genetic distance of *D. edmondsii* (BOLD accession GEONC001-22) was 10.3–10.5% (K2P) with *D. natalata* and 11.0–11.5% with *D. stellata*, while the distance between the latter two was 6.2–7.1%. The alignment was suitable for phylogenetic analysis, as no evidence of stop codons was detected and the index of substitution saturation was smaller than the critical value (ISS < ISS.C; p < 0.001) in the Xia test. The ML analysis (Fig. 4) clustered (*D. edmondsii* (*D. natalata* + *D. stellata*)) with high support. Although each genus had reasonable statistical support in the ML analysis, relationships between genera were not resolved.

Discussion

Although the identification of synapomorphies for *Disclisioprocta* deserves further assessments based on better knowledge of the morphology of related genera, the morphological similarities between the genitalia of *D. edmondsii* (Fig. 3) and *D. stellata* (Fig. 5) provide support to consider them as congenerics. The two species have uncus bifid, costal sclerotised band not reaching apex of the cucullus, a stout sacculus projection and a plate-like cornutus in the male and a flattened elongated antrum and a cluster of elongated spine-like signa arising from a plate on the corpus bursae in the female. The same morphological characters support the removal of *D. edmondsii* from *Xanthorhoe*. The type species of this genus has an elongated rod-like uncus, costa as a sclerotised free arm

extending beyond the cucullus apex, sacculus lacking a projection and numerous spinelike cornuti in the male and a short antrum and two elongated plate-like signa with small spines in the female (Expósito-Hermosa and Viidalepp 2011).

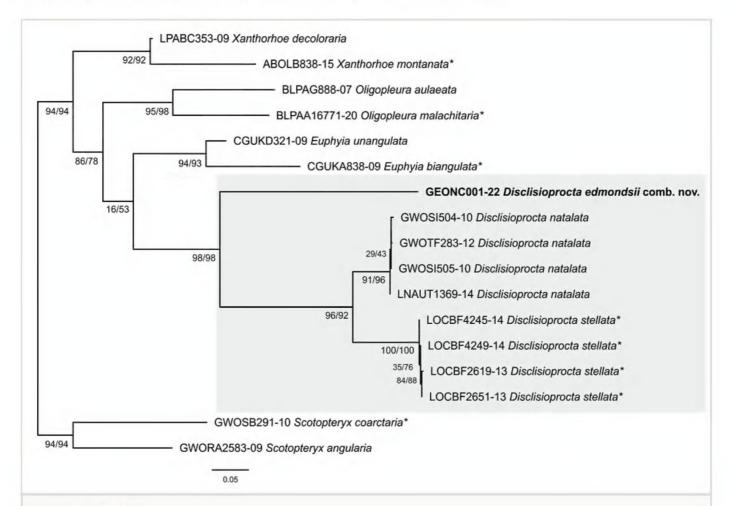


Figure 4. doi

Maximum Likelihood tree of *Disclisioprocta edmondsii* (Butler, 1882) comb. nov. (bold) and representatives of the Euphyiini + Xanthorhoini complex of Larentiinae, based on mitochondrial DNA sequences. Grey rectangle indicates *Disclisioprocta* Wallengren, 1861. Asterisks indicate type species. Numbers indicate SH-aLRT/UFBoot values (1000 replicates).

The result of the ML analysis is congruent with the genitalia morphology, since *D. edmondsii* clustered with two other representatives of *Disclisioprocta* in a well-supported clade, distantly related to members of *Xanthorhoe* (Fig. 4). The grouping of *D. stellata* and *D. natalata* as sister species agrees with the already highlighted remarkable morphological similarity of their genitalia (Hausmann 2009). The genetic distances of *D. edmondsii* with *D. natalata* and *D. stellata* seem higher than commonly reported for other Larentiinae genera (Stadie et al. 2014, Wanke et al. 2019, Guerrero et al. 2022). However, sequences deposited in BOLD (Ratnasingham and Hebert 2007) suggest that *Disclisioprocta* harbours several more species than the three recognised until recently (Parsons et al. 1999). Accordingly, it is highly probable that, with an improvement in the knowledge of the taxonomic diversity of *Disclisioprocta*, the genetic distance of *D. edmondsii* to other congenerics would be smaller. Alternatively, further studies could reveal *D. edmondsii* as a member of another, either described or undescribed, lineage of generic level, sister to *Disclisioprocta*. In the meantime, its placement in this genus seems a better solution than its previous adscription to *Xanthorhoe*, in spite the deep genetic distance with congenerics.

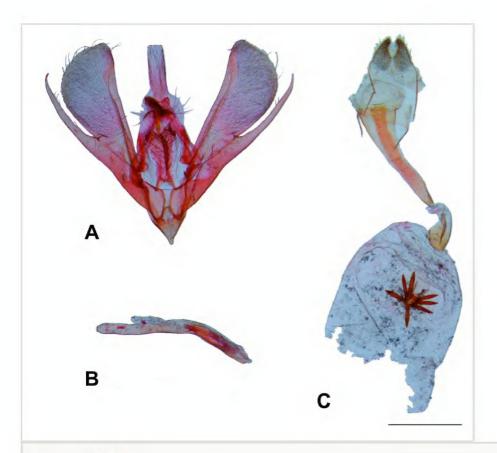


Figure 5. doi

Genitalia of *Disclisioprocta stellata* (Guenée, [1858]) from Brazil. **A** Male genitalia, phallus removed; **B** Phallus; **C** Female genitalia. Scale bar 1 mm.

Recent molecular phylogenetic analyses clustered *Disclisioprocta* with the Neotropical *Ptychorrhoe* Warren, 1900 in a clade sister to Euphylini (Brehm et al. 2019, Murillo-Ramos et al. 2019). Unfortunately, the genitalia of *Ptychorrhoe* remain unknown, impeding comparisons with *D. edmondsii*. Further morphological and molecular phylogenetic studies involving members of the Euphylini + Xanthorhoini complex (Brehm et al. 2019) are encouraged to determine the circumscription of the genera, identifying their synapomorpies and improving the current understanding of the evolution of Larentiinae.

Acknowledgements

I would like to thank Axel Hausmann for providing helpful comments and suggestions on a previous version of the manuscript, Olaf H. H. Mielke for the loan of the *R. affirmata* abdomens for dissection, Luis E. Parra for sharing a photo of the type material of *D. edmondsii* and Lafayette Eaton for checking the English.

References

- Angulo AO, Casanueva ME (1981) Catálogo de los Lepidópteros geométridos de Chile (Lepidoptera: Geometridae). Boletín de la Sociedad de Biología de Concepción 51: 7-39.
- Brehm G, Hebert PN, Colwell R, Adams M, Bodner F, Friedemann K, Möckel L, Fiedler K (2016) Turning up the heat on a hotspot: DNA barcodes reveal 80% more species of

- geometrid moths along an Andean Elevational Gradient. PLOS One 11 (3): e0150327. https://doi.org/10.1371/journal.pone.0150327
- Brehm G (2018) Revision of the genus Callipia Guenée, 1858 (Lepidoptera,
 Geometridae), with the description of 15 new taxa. European Journal of Taxonomy 404:
 1-54. https://doi.org/10.5852/ejt.2018.404
- Brehm G, Murillo-Ramos L, Sihvonen P, Hausmann A, Schmidt BC, Õunap E, Moser A, Mörtter R, Bolt D, Bodner F, Lindt A, Parra LE, Wahlberg N (2019) New World geometrid moths (Lepidoptera: Geometridae): Molecular phylogeny, biogeography, taxonomic updates and description of 11 new tribes. Arthropod Systematics & Phylogeny 77: 457-486. https://doi.org/10.26049/ASP77-3-2019-5
- Butler AG (1882) Heterocerous Lepidoptera collected in Chile by Thomas Edmond Esq. Part. III-Geometrites. Transactions of the Entomological Society of London 30: 339-427. https://doi.org/10.1111/j.1365-2311.1882.tb01580.x
- Chalup A (2014) Geometridae. In: Roig-Juñent S, Claps LE, Morrone JJ (Eds)
 Biodiversidad de Artrópodos Argentinos Volumen 4. INSUE UNT, San Miguel de
 Tucumán, 547 pp.
- Expósito-Hermosa A, Viidalepp J (2011) Xanthorhoe iberica (Staudinger, 1901) sp. bon., de España (Lepidoptera: Geometridae, Larentiinae, Xanthorhoeini). SHILAP Revista de Lepidopterología 39 (156): 419-422.
- Folmer O, Black M, Hoeh W, Lutz R, Vrijenhoek R (1994) DNA primers for amplification of mitochondrial cytochrome c oxidase subunit I from diverse metazoan invertebrates.
 Molecular Marine Biology and Biotechnology 3: 294-299.
- Guerrero JJ, Hausmann A, Rubio RM, Garre M, Ortiz AS (2022) First description of the male and DNA barcode of *Euphyia vallantinaria* (Oberthür, 1890) from the Iberian Peninsula (Lepidoptera, Geometridae, Larentiinae). Nota Lepidopterologica 45: 33-39. https://doi.org/10.3897/nl.45.75693
- Guindon S, Dufayard J, Lefort V, Anisimova M, Hordijk W, Gascuel O (2010) New algorithms and methods to estimate maximum-likelihood phylogenies: Assessing the performance of PhyML 3.0. Systematic Biology 59 (3): 307-321. https://doi.org/10.1093/sysbio/syg010
- Hausmann A (2009) New and interesting geometrid moths from Sokotra islands (Lepidoptera, Geometridae). Mitteilungen der Münchner Entomologischen Gesellschaft 99: 95-104.
- Hausmann A, Parra LE (2009) An unexpected hotspot of moth biodiversity in Chilean northern Patagonia (Lepidoptera, Geometridae). Zootaxa 1989: 23-38. https://doi.org/10.11646/zootaxa.1989.1
- Hebert PN, Cywinska A, Ball S, deWaard J (2003) Biological identifications through DNA barcodes. Proceedings of the Royal Society of London. Series B: Biological Sciences 270 (1512): 313-321. https://doi.org/10.1098/rspb.2002.2218
- Hoang DT, Chernomor O, von Haeseler A, Minh BQ, Vinh LS (2017) UFBoot2: Improving the Ultrafast Bootstrap Approximation. Molecular Biology and Evolution 35 (2): 518-522. https://doi.org/10.1093/molbev/msx281
- Kalyaanamoorthy S, Minh BQ, Wong TKF, von Haeseler A, Jermiin LS (2017)
 ModelFinder: fast model selection for accurate phylogenetic estimates. Nature Methods 14 (6): 587-589. https://doi.org/10.1038/nmeth.4285

 Matson TA (2022) A new monotypic genus from the American southwest to accommodate "Semiothisa" kuschea (Geometridae: Ennominae). Zootaxa 5093: 067-074. https://doi.org/10.11646/zootaxa.5093.1.4

- Moraes S, Montebello Y, Stanton M, Yamaguchi LF, Kato M, Freitas AVL (2021)
 Description of three new species of Geometridae (Lepidoptera) using species delimitation in an integrative taxonomy approach for a cryptic species complex. PeerJ 9: e11304. https://doi.org/10.7717/peerj.11304
- Murillo-Ramos L, Sihvonen P, Brehm G, Ríos-Malaver IC, Wahlberg N (2021) A
 database and checklist of geometrid moths (Lepidoptera) from Colombia. Biodiversity
 Data Journal 9: e68693. https://doi.org/10.3897/BDJ.9.e68693
- Nguyen L, Schmidt H, von Haeseler A, Minh BQ (2014) IQ-TREE: A fast and effective stochastic algorithm for estimating maximum-likelihood phylogenies. Molecular Biology and Evolution 32 (1): 268-274. https://doi.org/10.1093/molbev/msu300
- Parra LE (2018) A new genus of Nacophorini (Geometridae) from Chile. Journal of the Lepidopterist's Society 72: 265-269. https://doi.org/10.18473/lepi.72i4.a3
- Parsons MS, Scoble MJ, Honey MR, Pitkin LM, Pitkin BR (1999) The Catalogue. In: Scoble MJ (Ed.) Geometrid Moths of the World: a Catalogue (Lepidoptera: Geometridae). CSIRO Publishing, Collingwood, 1016 pp. https://doi.org/10.1071/9780643101050
- Pitkin LM (2002) Neotropical ennomine moths: a review of the genera (Lepidoptera: Geometridae). Zoological Journal of the Linnean Society 135: 121-401. https://doi.org/10.1046/j.1096-3642.2002.00012.x
- Rambaut A (2014) FigTree, a graphical viewer of phylogenetic trees. Version 1.4.2.
 Institute of Evolotionary Biology, University of Edinburgh. URL: http://tree.bio.ed.ac.uk/software/figtree/
- Ramos-González M, Zamora-Manzur C, Saladrigas Menés D, Parra L (2019) The Trichopterygini (Lepidoptera, Geometridae) of Austral South America: description of new species from Chile. ZooKeys 832: 91-111. https://doi.org/10.3897/zookeys.832.30851
- Ratnasingham S, Hebert PDN (2007) BOLD: the barcode of life data system (www.barcodinglife.org). Molecular Ecology Notes 7: 355-367. https://doi.org/10.1111/j.1471-8286.2007.01678.x
- Stadie D, Hausmann A, Rajaei H (2014) Cataclysme subtilisparsata Wehrli, 1932 (Lepidoptera, Geometridae, Larentiinae) recognized as bona species - an integrative approach. Nota Lepidopterologica 37 (2): 141-150. https://doi.org/10.15468/kdnpwf
- Tamura K, Stecher G, Kumar S (2021) MEGA11: Molecular Evolutionary Genetics
 Analysis Version 11. Molecular Biology and Evolution 38 (7): 3022-3027. https://doi.org/10.1093/molbev/msab120
- Trifinopoulos J, Nguyen L, von Haeseler A, Minh BQ (2016) W-IQ-TREE: a fast online phylogenetic tool for maximum likelihood analysis. Nucleic Acids Research 44: W232-W235. https://doi.org/10.1093/nar/gkw256

- Vargas HA, Mielke OHH, Casagrande MM, Parra LE (2010) Imaturos de *Chrismopteryx* undularia (Blanchard) (Lepidoptera, Geometridae). Revista Brasileira de Entomologia 54 (4): 519-528. https://doi.org/10.1590/S0085-56262010000400001
- Vargas HA (2021) A new species of Eupithecia Curtis (Lepidoptera, Geometridae) from the Andes of northern Chile. Nota Lepidopterologica 44: 239-247. https://doi.org/10.3897/nl.44.73247
- Vargas HA, Solis MA, Vargas-Ortiz M (2022) The South American moth Rheumaptera mochica (Dognin, 1904) (Lepidoptera, Geometridae, Larentiinae) rediscovered after more than a century of anonymity. ZooKeys 1085: 129-143. https://doi.org/10.3897/zookeys.1085.76868
- Viidalepp J (2011) A morphological review of tribes in Larentiinae (Lepidoptera: Geometridae). Zootaxa 3136: 1-44. https://doi.org/10.11646/zootaxa.3136.1.1
- Viidalepp J, Lindt A (2019) A new Neotropical emerald moth genus based on some unusual "artefacts" (Lepidoptera: Geometridae, Geometrinae). Zootaxa 4691 (2): 181-187. https://doi.org/10.11646/zootaxa.4691.2.8
- Wanke D, Hausmann A, Rajaei H (2019) An integrative taxonomic revision of the genus Triphosa Stephens, 1829 (Geometridae: Larentiinae) in the Middle East and Central Asia, with description of two new species. Zootaxa 4603 (1). https://doi.org/10.11646/2001axa.4603.1.2
- Wanke D, Feizpour S, Hausmann A, Viidalepp J, Rajaei H (2022) Taxonomy and systematics of the enigmatic emerald moth *Xenochlorodes graminaria* (Kollar, 1850) (Lepidoptera: Geometridae), and its assignment to a new genus. Integrative Systematics: Stuttgart Contributions to Natural History 5 (1): 61-71. https://doi.org/10.18476/2022.857803
- Xia X (2018) DAMBE7: New and improved tools for data analysis in molecular biology and evolution. Molecular Biology and Evolution 35 (6): 1550-1552. https://doi.org/10.1093/molbev/msy073